# JOURNAL OF Evolutionary Biology

.04ese

# Does increased heat resistance result in higher susceptibility to predation? A test using *Drosophila melanogaster* selection and hardening

S. HANGARTNER\*† (D, I. DWORKIN<sup>‡</sup>, M. DENIEU§ & A. A. HOFFMANN<sup>†</sup>

\*School of Biological Sciences, Monash University, Clayton, Vic., Australia

†School of BioSciences, University of Melbourne, Bio21 Institute, Parkville, Vic., Australia

Department of Biology, McMaster University, Hamilton, ON, Canada

§Department of Biological Sciences, George Washington University, Washington, DC, USA

#### Keywords:

biotic interactions; climate change; genetic correlations; phenotypic plasticity; thermal adaptation; trade-offs.

### Abstract

Heat resistance of ectotherms can be increased both by plasticity and evolution, but these effects may have trade-offs resulting from biotic interactions. Here, we test for predation costs in Drosophila melanogaster populations with altered heat resistance produced by adult hardening and directional selection for increased heat resistance. In addition, we also tested for genetic tradeoffs by testing heat resistance in lines that have evolved under increased predation risk. We show that while 35/37 °C hardening increases heat resistance as expected, it does not increase predation risk from jumping spiders or mantids; in fact, there was an indication that survival may have increased under predation following a triple 37 °C compared to a single 35 °C hardening treatment. Flies that survived a 39 °C selection cycle showed lower survival under predation, suggesting a predation cost of exposure to a more severe heat stress. There was, however, no correlated response to selection because survival did not differ between control and selected lines after selection was relaxed for one or two generations. In addition, lines selected for increased predation risk did not differ in heat resistance. Our findings suggest independent evolutionary responses to predation and heat as measured in laboratory assays, and no costs of heat hardening on susceptibility to predation.

### Introduction

Resistance to thermal extremes including high temperatures is an important factor influencing the distribution and abundance of animal species (Cossins & Bowler, 1987; Hoffmann *et al.*, 2013). As high temperature stresses are expected to become more common in severity and frequency, species are increasingly at risk of exposure to conditions exceeding their upper thermal limit (IPCC 2014). However, upper thermal limits of ectotherms can be increased to some extent by both plastic and evolutionary responses (Hoffmann & Parsons, 1991; Angilletta, 2009; Hoffmann *et al.*, 2013).

*Correspondence:* Sandra Hangartner, School of Biological Sciences, Monash University, Clayton 3800, Vic., Australia. Tel.: (03) 9905 2718; e-mail: sandra.hangartner@monash.edu

Plastic responses are rapid, occur within an organism's lifetime and have been predicted to play a major role in thermal adaptation (Charmantier et al., 2008; Chevin et al., 2013). In particular, many studies have examined the benefits of hardening responses to heat exposure, whereby heat resistance is enhanced by prior exposure to a moderate heat stress (Hoffmann et al., 2003; Sinclair et al., 2003; Angilletta, 2009). Moreover, plasticity may itself evolve in response to environmental change (Scheiner, 1993; Kingsolver et al., 2007; but see Sørensen et al., 2016). In addition to plastic responses, upper thermal limits can be altered by selection, resulting in populations that differ in levels of resistance to heat stress (Hoffmann et al., 2013). Selection responses have been particularly well studied in Drosophila melanogaster, where artificial selection can increase resistance to heat stress (McColl et al., 1996; Gilchrist & Huey, 1999; Bubliy & Loeschcke, 2005;

Hangartner & Hoffmann, 2016) and populations can be differentiated along climatic gradients as well (Hoffmann *et al.*, 2002). These results demonstrate that standing genetic variation and new mutations are sufficient to drive the evolutionary response. However, evolutionary changes in heat responses might nevertheless be limited (Schou *et al.*, 2014; Hangartner & Hoffmann, 2016).

Although plastic and evolutionary changes can allow insects to adapt to stressful, hot conditions, their benefits are likely to be curtailed by costs of plastic and genetic shifts. Hardening responses may incur costs that become evident in terms of growth rates (e.g. Feder et al., 1992), longevity (e.g. Bubliy et al., 2012, 2013) or fecundity (e.g. Krebs & Loeschcke, 1994; Hercus et al., 2003; Huang et al., 2007). However, the costs and benefits of hardening have been defined mainly in laboratory assays of fitness, making the ecological significance of hardening unclear (Loeschcke & Hoffmann, 2007). Field release studies have revealed costs associated with acclimation or hardening that laboratory-based assays did not detect, highlighting the importance of integrating ecological parameters that affect costs and benefits of phenotypic plasticity (Loeschcke & Hoffmann, 2007; Kristensen et al., 2008). In addition, genetic constraints may act to limit evolutionary responses. Genetically correlated traits do not evolve independently, and the covariances between traits can either facilitate or hamper adaptation (Walsh & Blows, 2009).

To date, costs associated with heat resistance have mostly been considered through potential trade-offs with performance measures under favourable conditions or under opposing thermal extremes (e.g. Huev & Kingsolver, 1993; Willett, 2010; Karl et al., 2014). On the other hand, biotic interactions around predation, parasitism and competition can also form an important component of resistance costs. Because species interactions are expected to significantly alter climate change responses, interspecific relationships should be incorporated into the predictive framework of climate change (Sanford, 1999; Harley, 2011; Miller et al., 2014). The effects of heat stressors may be magnified by biotic stressors such as predators (e.g. Relyea & Mills, 2001; Alton et al., 2010). Most natural populations are likely to experience selection from multiple abiotic and biotic selective pressures concurrently; however, our understanding of adaptation to multiple selective agents is still limited (but see Ghalambor et al., 2004; Schulte, 2007; Eränen et al., 2009; Rogell et al., 2009; Egea-Serrano et al., 2014).

Predation is an important selective force in natural systems and can impose strong selection on antipredator traits. Adaptive responses to predation risk may be modified by other stressors, as environmental stress can increase the costs (Hanazato, 2001; Huber *et al.*, 2004; Teplitsky *et al.*, 2005) or decrease investment in defences (Barry, 2000; Relyea, 2004; Teplitsky *et al.*,

2007). A few studies have tested for heritability in predator avoidance traits and usually found substantial genetic variation (Grant & Mettler, 1969; Stirling et al., 2002; Relyea, 2005; Brokordt et al., 2012; DeNieu et al., 2014; but see Blumstein et al., 2010). Whether predator avoidance is genetically correlated with other types of stress resistance is largely unknown (but see Jansen et al., 2011). Predation and heat stress resistance might influence the same behavioural or physiological aspects of an organism's biology (Miller et al., 2014). Exposure to heat stress has, for example, been shown to reduce metabolic rates (Dinh et al., 2016), which potentially could affect survival under predation risk (Rovero et al., 1999; Beckerman et al., 2007; Slos & Stoks, 2008). Whether predation pressure alters the costs and benefits of heat hardening and evolved heat resistance has never been tested.

We first tested for the ability of flies to avoid predators in D. melanogaster populations with altered heat resistance produced by adult hardening and directional selection for heat resistance. We measured the costs and benefits of single and repeated heat hardening treatments by subjecting flies that were exposed to different hardening treatments to predation by juvenile false garden mantids (Pseudomantids albofimbriata) and jumping spiders (Salticidae spp.). Both predators have excellent eyesight, slowly approach their prey and capture them with a rapid movement (Gelperin, 1968; Jackson & Pollard, 1996). Costs (or benefits) of the hardening treatments would be evidenced by increased (or decreased) survival of flies from the hardening treatments compared to controls. Differential survival among the hardening treatments would suggest intensity- and/or frequency-dependent costs (or benefits) under predation pressure.

Second, lines that have been selected for increased heat resistance were tested for performance under predation by the same two predators. Experiments were performed after zero, one and two generations of relaxed selection to test for costs of stress exposure and genetic trade-offs. In addition, we also tested for genetic trade-offs between heat resistance and predator avoidance using lines that have evolved under predation risk by juvenile Chinese mantids (Tenodera aridifolia sinensis) or zebra jumping spiders (Salticus scenicus). These lines were tested for heat resistance after two generations of relaxed selection. Genetic trade-offs between heat resistance and predator avoidance would be evident if heatresistant lines have lower survival under predation than control lines after two generations without selection (generation 2). In addition, lines evolved under predation risk would be expected to be less heat resistant than control lines after two generations of relaxed selection. Costs (or benefits) of a more severe stress exposure would be evident as stress-resistant lines having lower (or higher) survival after the selection cycle (generation 0). Reduced (or increased) survival of heat-

### **Materials and methods**

#### Fly cultures

All cultures were held at constant 19 °C, under 12: 12-h light: dark cycle in 250-ml bottles containing laboratory medium composed of dextrose (7.5% w/v), cornmeal (7.3% w/v), inactive yeast (3.5% w/v), soya flour (2% w/v), agar (0.6% w/v), 4-methyl 4-hydroxybenzo-ate (1.6%) and acid mix (1.4% 10:1 propionic acid: orthophosphoric acid). The experimental flies were reared under controlled density conditions by removing parents from the bottles after 48 h of oviposition.

# Testing predator avoidance after heat hardening treatments

### Heat hardening treatments

The experimental flies originated from a mass-bred population that was collected near Melbourne in May 2012 and was maintained under standard laboratory conditions as described above. Two- to 3-day-old flies were separated by sex under light CO<sub>2</sub> anaesthesia on 17 December 2013 and held in separate vials according to sex, at a density of 25 individuals per vial (day 1). The hardening experiments were started at 4–5 days posteclosion, and flies were randomly allocated to the five heat hardening treatments. Females and males were kept separately throughout the hardening experiments, which enabled us to test for sex effects of the hardening treatments on heat resistance.

For the heat hardening, glass bottles (100 mL) containing 50 females or males were immersed in a circulating water bath at either 35 or 37 °C. Temperature was controlled using a Ratek SP599 thermoregulator with a REXP24 controller (Ratek, Boronia, Vic, Australia). In the control treatment, flies were kept in bottles for 75 min at 19 °C on day 3, day 5 and day 7. Flies in the single 35 °C (35-1) treatment received one hardening treatment of 75 min at 35 °C on day 7. Flies in triple 35 °C (35-3) treatment received three hardening treatments of 75 min at 35 °C on day 3, day 5 and day 7. The single 37 °C (37-1) treatment consisted of one hardening treatment of 75 min at 37 °C on day 7. Finally, the triple 37 °C (37-3) treatment involved three hardening treatments of 75 min at 37 °C on day 3, day 5 and day 7.

### *Heat resistance experiments*

Flies were tested for heat resistance after the heat hardening treatments to test whether the hardening increased resistance. Ten females and males per treatment were tested for heat resistance at static 39.0 °C. These experiments were performed on day 8 in two blocks, where five females and males per treatment were tested in each block. To score heat resistance, flies were placed individually into 5-mL vials submerged into a glass tank with water held at 39.0 °C. Each fly was scored for heat resistance, where resistance was defined as the time taken for each fly to be knocked down and become immobile even when exposed to a flashlight.

#### Predator cultures

Female adult false garden mantids (Pseudomantids albofimbriata) and mantid egg cases were collected between March and May 2013 and juvenile jumping spiders (Salticidae spp.) in June and July 2013 near Melbourne. Juvenile jumping spiders were kept individually in vials. Although it was not possible to identify spiders to species, we exposed all treatments to the spider simultaneously to ensure that any species differences did not confound the detection of treatment effects. Female adult mantids were kept individually in containers where they laid egg cases. The egg cases were hatched and maintained at 19 °C. The hatching mantids were collected and kept individually in vials containing fly medium and Drosophila as a food source. Predators were kept at constant 19 °C, under 12: 12-h light: dark cycles. All animals were fed on Drosophila, and vials or containers had fly medium and twigs as a substrate for the spiders and mantids.

#### Predation experiments

We tested for survival of the flies that had been exposed to heat hardening treatments under predation by jumping spider (Salticidae spp.) and juvenile mantids (Pseudomantids albofimbriata). Flies were exposed to predators after a recovery period of 8 h after the hardening treatment on day 7. These experiments were performed separately for the two predators, and females and males were tested in separate vials/bottles. Flies originating from different treatments were colourmarked by lightly shaking them in a vial containing micronized fluorescent dust (Radiant). Five different colour combinations were used to test for any potential effects of a particular colour, whereas each treatment had a different colour in each colour combination. One fly per treatment (total of five flies) were exposed to one spider in a vial  $(28 \times 95 \text{ mm})$  containing laboratory medium as food for the flies and some branches which provided structural complexity and shelter for the flies and spiders. Each of the five colour combination was replicated eight times resulting in a total of 40 replicates (vials) for both sexes (200 females and males in total) for the spider experiments. For the mantids, the experimental procedure was similar; however, two flies per treatment (total of 10 flies) were exposed to one mantid in a bottle ( $6 \times 13$  cm). Each of the five

colour combinations was replicated four times resulting in a total of 20 replicates (bottles) for both sexes (200 females and males in total). Surviving flies were removed from the vial/bottle when about 50% of the flies have been predated (after 1–5 days) and survived flies were scored for treatment origin using the colour markings. No natural mortality was observed during the experiments. Survival (yes or no) of each fly was used for the statistical analyses.

# Testing heat resistance selection lines for predation avoidance

#### *Heat resistance selection lines*

The heat-selected lines have been described in detail in Hangartner & Hoffmann (2016). In short, all selected and control lines were founded from D. melanogaster collected near Melbourne in May 2012. The offspring of 60 field-collected females were pooled and mass-bred for two generations in the laboratory prior to the first selection at generation F3 for the heat-resistant selected lines. The selection experiments were carried out separately for both sexes, and the top 10% most resistant flies were selected and randomly allocated into five replicate lines per selection regime comprised of 90-110 flies of each sex (200-210 in total). Flies were selected for heat knockdown resistance by immersing glass bottles (100 mL) containing 100 flies in a circulating water bath at 39 °C. When ca 90% of the flies were knocked down (did not move anymore when flashed with a flash light), bottles were removed from the tank and the remaining 10% of flies that were able to stand up were selected (for further details, see Hangartner & Hoffmann, 2016). The control lines were established and maintained in the same manner as the heat-resistant lines, but these lines were not exposed to any treatment. The heat-resistant lines have evolved to have a tolerance level around 0.5 °C higher than the control lines after ten generations of strong selection (Hangartner & Hoffmann, 2016).

#### Predation experiments

We scored the heat-selection lines for survival under predation by jumping spiders (*Salticidae* spp.) and juvenile mantids (*Pseudomantids albofimbriata*) to test for costs or benefits of a severe stress exposure and genetic trade-offs. These were performed on adult flies after one and two generations of relaxed selection, as well as right after the selection experiment (no relaxation). The experiments were performed separately for the two predators, and the sexes were tested in separate vials/bottles. Flies were between 4 and 7 days old at the beginning of the experiment. However, selected flies were slightly older (9–12 days), as selection experiments were performed on them before. The control flies had the same age as the selected flies, which means that any potential age effect of the flies would apply to both, control and selected flies.

Six adult flies were randomly chosen from three different control and selected lines for the spider experiments, whereas 12 adults were chosen for the mantids experiments. Flies were marked with dust colours as described above, where each fly (line) was assigned a colour. Ten different colour combinations were used to account for potential colour effects. Each colour combination was replicated five times for the spider experiments resulting in a total of 50 replicates (vials) for both sexes (300 flies in total per sex). For the mantid experiments, each colour combination was replicated three times resulting in a total of 30 replicates per sex (360 flies in total per sex). The colour-marked flies were exposed to one spider or one mantid in a vial  $(28 \times 95 \text{ mm})$  or bottle  $(6 \times 13 \text{ cm})$ , respectively, containing fly food and some twigs which provided structural complexity and shelter for the flies and predators. Surviving flies were removed from the vial/bottle when about half of the flies have been predated, and scored for the line origin based on colour markings. Survival (yes or no) of each fly was used for the statistical analyses.

#### Testing predation selection lines for heat resistance

In addition, we also tested for genetic trade-offs between heat resistance and predator avoidance by scoring heat resistance in lines that have evolved under predation risk by jumping spiders or mantids. These flies were derived from the Dworkin Laboratory at Michigan State University, USA. Two sets of selection lines were used (episodic and continuous predation), which are described in detail in the Data S1. The episodic and continuous predation lines differed in effective population size and the strength of selection induced by the predators. Including both sets of selection lines allowed us to test whether effective population size and/or strength of selection may affect the detection of an apparent trade-off. The episodic and continuous predation lines were tested for heat resistance after two generations without selection to ensure that any differences found in the subsequent experiments were genetic rather than due to plastic (cross-generation) effects (c.f. Schiffer et al., 2013). Ten females and males per line were tested for heat resistance at static 39.0 °C. These experiments were performed separately for the episodic and continuous predation regimes and in two blocks per regime, where five females and males per line were tested in each block. Flies were sexed under light CO<sub>2</sub> anaesthesia and scored for heat resistance when they were 4-5 days old. To score heat resistance, flies were placed individually into 5-mL vials submerged into a glass tank with water held at 39.0 °C. Each fly was scored for heat tolerance, where tolerance was defined as the time taken for each fly to be

knocked down and become immobile even when exposed to a flashlight.

#### Statistical analyses

All statistical analyses were performed with SAS 9.3 (SAS Institute, Inc., Cary, North Carolina, USA) and involved general and generalized linear models. *Post hoc* pairwise comparisons were undertaken using Tukey's tests comparing least square means and adjusting for multiple comparisons.

Heat resistance of the hardened flies was analysed using a general linear model with the GLM procedure and Kenward–Roger degrees of freedom method (Littell *et al.*, 2006). Hardening treatment, sex, the hardening treatment  $\times$  sex interaction and block were included as fixed factors in this analysis.

Survival under predation after the hardening treatments was analysed with a generalized linear mixed model with REML estimation, logit link function and a binary distribution using the Proc GLIMMIX. REML specification performs residual (restricted) maximum likelihood, where negative estimates are constrained to zero (Littell *et al.*, 2006). In these analyses, sex, treatment and their interaction were included as fixed factors. To test for any potential effect of a particular colour on survival, colour was included as an additional fixed factor. In addition, vial (nested under sex) was included as a random factor.

Survival under predation of the selection lines was analysed with a generalized linear mixed model with REML estimation, logit link function and a binary distribution using the Proc GLIMMIX (Littell *et al.*, 2006). Separate models were run for each generation (zero, one or two generations of relaxation). In these models, selection regime and sex as well as their interactions were included as fixed factors. In addition, colour was included as a fixed factor and vial (nested within sex) and line (nested within selection regime) were included as random factors.

Heat knockdown time of the predation selection lines was normally distributed and was analysed using linear mixed model analyses of variance with the MIXED procedure and Kenward–Roger degrees of freedom method (Littell *et al.*, 2006). Selection regime and sex were included as fixed factors and line (nested under selection regime) as a random factor.

### Results

## Testing predator avoidance after heat hardening treatments

#### Hardening effects on heat knockdown time

The hardening treatments had a significant effect on heat resistance, with all hardening treatments increasing heat knockdown time of the flies by 21–73%

**Table 1** General linear model of heat knockdown time (min) of

 female and male *D. melanogaster* after five different heat hardening

 treatments. Eta square is the proportion of total variation

 accounted for by the effect being tested. Significant values are

 shown in bold.

Effect	d.f.	Eta square	Mean square	F	Р
Block	1	0.001	7.14	0.16	0.686
Sex	1	0.006	46.03	1.06	0.305
Treatment	4	0.213	434.58	10.01	< 0.001
$Sex \times treatment$	4	0.041	83.36	1.92	0.110



**Fig. 1** Mean  $\pm$  SE heat knockdown time (min) of female and male *D. melanogaster* after a control treatment, a single 35 °C (35-1) and 37 °C (37-1) hardening treatment and a triple 35 °C (35-3) and 37 °C (37-3) hardening treatment.

compared to the control treatment (Table 1, Fig. 1). *Post hoc* Tukey's tests revealed that all hardening treatments significantly increased heat resistance (not shown). In addition, flies from the triple 37 °C hardening treatment had a significantly higher heat resistance than flies from the triple 35 °C hardening treatment at  $t_{139} = 2.89$  (adjusted P = 0.004), increasing heat resistance by about 30% (Fig. 1). The sexes did not significantly differ in heat resistance, and the sex × treatment interaction was not significant (Table 1, Fig. 1).

#### *Predation avoidance after heat hardening*

The hardening treatments had a significant effect on survival under predation by jumping spiders (Table 2A, Fig. 2a). A *post hoc* Tukey's test revealed that flies from the triple 37 °C hardening treatment had significantly

© 2017 EUROPEAN SOCIETY FOR EVOLUTIONARY BIOLOGY. J. EVOL. BIOL. 30 (2017) 1153–1164 JOURNAL OF EVOLUTIONARY BIOLOGY © 2017 EUROPEAN SOCIETY FOR EVOLUTIONARY BIOLOGY

**Table 2** Generalized linear mixed model of binary survival (yes or no) of female and male *D. melanogaster* under predation pressure by (A) jumping spiders and (B) juvenile mantids. Flies originating from five different heat hardening treatments. For the fixed effects, ndf is numerator degrees of freedom, and ddf is

denominator degrees of freedom. For the random effects, var is the random effects variance component. Significant values are shown in bold.

		(A) Spiders				(B) J	(B) Juvenile mantis			
Effect		ndf	ddf	F	Р	ndf	ddf	F	Ρ	
Fixed factors										
Sex		1	57	0.2	0.660	1	28	1.6	0.220	
Treatment		4	224	2.5	0.043	4	254	0.1	0.976	
Sex $\times$ treat	tment	4	224	0.9	0.494	4	254	3.3	0.012	
Colour		4	224	1.1	0.375	4	254	0.7	0.596	
Effect	var		SE	Ζ	Ρ	var	SE	Z	Г Р	
Random facto Vial (sex)	ors 0.00	1				0.00				

higher survival than those from the single 35 °C hardening treatment at  $t_{224} = 3.01$  (adjusted P = 0.024), with a survival difference of around 29%. The sexes did not differ in survival, and the treatment × sex interaction was not significant. In addition, colour did not have a significant effect on survival (Table 2A). Heat hardening treatments, sex and colour did not have a significant effect on survival under predation by juvenile mantids, but the treatment × sex interaction was significant (Table 2B, Fig. 1b). A *post hoc* Tukey's test revealed that the difference between the sexes in the single 37 °C hardening treatment was observed nonsignificant ( $t_{254} = 318$ , adjusted P = 0.051) (Fig. 2b).

# Testing heat resistance selection lines for predation avoidance

Lines that have been selected for increased heat resistance were tested for predation avoidance immediately after selection (generation 0), as well as after one and two generations without selection.

Generation 0: The analysis at generation 0 showed that the selection regime had a significant effect on survival under predation risk by spiders and mantids: heat-resistant lines had significantly lower survival than control lines after the selection cycle reflecting an average survival difference of 10% under predation by spiders and 17% under predation by mantids (Table 3, Fig. 3). Survival under predation by spiders did not significantly differ between sexes, and the selection regime  $\times$  sex interaction was not significant (Table 3A). The sexes did not differ for survival under predation by mantids, but there was a significant selection regime  $\times$  sex interaction. Post hoc tests revealed that the selection regimes did not significantly differ for the females ( $t_{597} = 0.97$ , P = 0.331), but there was a significant difference for the males  $(t_{597} = 3.75, P < 0.001)$ , where survival of the control males was 25% higher than the survival of the selected males (Fig. 3b).



**Fig. 2** Mean  $\pm$  SE survival (%) of female and male *D. melanogaster* after different heat hardening treatments when exposed to (a) jumping spiders and (b) juvenile mantids. Hardening treatments included a control treatment, a single 35 °C (35-1) and 37 °C (37-1) hardening treatment and a triple 35 °C (35-3) and 37 °C (37-3) hardening treatment.

**Table 3** Generalized linear mixed model of binary survival (yes or no) of female and male *D. melanogaster* under predation pressure by (A) jumping spiders and (B) juvenile mantids. Flies originated from control and heat-resistant selection lines, and the predation experiment was performed after the selection experiments (generation 0). For the fixed effects, ndf is numerator degrees of freedom, and ddf is denominator degrees of freedom. For the random effects, var is the random effects variance component. Significant values are shown in bold.

	(A) Spiders				(B) Juvenile mantis			
Effect	ndf	ddf	F	Р	ndf	ddf	F	Р
Fixed factors								
Selection regime	1	8	5.57	0.046	1	8	7.93	0.023
Sex	1	98	0.12	0.725	1	52	0.68	0.414
Selection regime × sex	1	476	0.47	0.493	1	574	5.11	0.024
Colour	5	476	0.77	0.570	5	574	1.19	0.313
Effect	var	SE	= <i>Z</i>	Ρ	var	ç	SE .	Z P
Random factors Vial (sex) Line (selection regime)	0.00 0.00				0.00 0.00	) . ) .		

*Generations 1 and 2:* After one and two generations without selection, there was no significant difference of survival between the selection regimes under both

predators (Tables S1 and S2, Fig. 4). The sexes and the sex  $\times$  selection regime interaction had no significant effect on survival under predation from either spiders or mantids at generations 1 and 2 (Tables S1 and S2, Fig. 4).

# Testing for heat resistance in predation selection lines

Next, we investigated heat resistance in lines that have evolved under predation risk with both jumping spiders and mantids. The selection regime as well as the selection regime  $\times$  sex interaction did not have a significant effect on heat knockdown time in both the episodic and continuous predation regimes (Table 4A,B). Heat knockdown time did, however, differ between the sexes in the episodic and continuous predation selection regimes, whereas males were more heat tolerant than females overall, where heat resistance of males was roughly 30% higher than female resistance (Table 4A, B). In addition, there was significant variation among the lines within the selection regimes in the continuous predation lines (Table 4B).

### **Discussion**

To our knowledge, this is the first study to consider both the impact of evolutionary changes in heat resistance and heat hardening on susceptibility to



At generation 0 (no relaxation)

**Fig. 3** Mean  $\pm$  SE survival (%) of female and male *D. melanogaster* under predation by (a) jumping spiders and (b) juvenile mantids of control lines (control) and lines selected for increased heat resistance (heat). Survival was scored at generation 0 (no relaxation).

© 2017 EUROPEAN SOCIETY FOR EVOLUTIONARY BIOLOGY. *J. EVOL. BIOL.* **30** (2017) 1153–1164 JOURNAL OF EVOLUTIONARY BIOLOGY © 2017 EUROPEAN SOCIETY FOR EVOLUTIONARY BIOLOGY



**Fig. 4** Mean  $\pm$  SE survival (%) of female and male *D. melanogaster* under predation by (a) jumping spiders and (b) juvenile mantids of control lines (control) and lines selected for increased heat resistance (heat). Survival was scored at generations 1 and 2 (1 and 2 generations of relaxation).

predation and the impact of selection for predation avoidance on heat resistance. We found that heat hardening with a higher frequency and intensity can increase survival under predation by spiders. However, there was no evidence for a genetic trade-off between heat resistance and predator avoidance because survival did not differ between control and heat-selected lines and lines selected for increased predation risk did not differ in heat resistance after selection was relaxed for two generations. Survival after the selection cycle at generation 0 was, however, reduced, which suggests that a severe heat stress can reduce survival under predation. Our finding that frequency- and intensitydependent heat exposure, but not evolutionary changes in heat resistance, affects predation avoidance is novel and has implications for taxa in the face of climate change.

# Costs and benefits of heat hardening under predation pressure

We did not find any costs of heat hardening under predation by either jumping spiders or juvenile mantids. We suspect that costs associated with hardening found in the field and leading to lower capture rates of released flies (Loeschcke & Hoffmann, 2007; Kristensen *et al.*, 2008) are unlikely to reflect increased susceptibility to predation. These results suggest that other costs of hardening are likely to be involved under field conditions.

Effect	(A) Episo	dic predation I	ines		(B) Continuous predation lines			
	ndf	ddf	F	Р	ndf	ddf	F	Р
Fixed factors								
Selection regime	1	2	0.30	0.638	2	9	0.10	0.908
Sex	1	74	25.40	< 0.001	1	223	37.81	< 0.0001
Selection regime $\times$ sex	1	74	2.26	0.137	2	223	1.77	0.172
Block					1	223	0.11	0.743
Effect	var	SE	Ζ	Р	var	SE	Ζ	Р
Random factors								
Line (selection regime)	0.99	5	0.21	0.417	24.2	14	1.73	0.042
Residual	74.42	12	6.08	< 0.001	110	10	10.56	< 0.001

**Table 4** Mixed model analyses of variance of heat knockdown time (min) of female and male *D. melanogaster*. Flies originating from the (A) episodic and (B) continuous predation lines. For the fixed effects, ndf is numerator degrees of freedom, and ddf is denominator degrees of freedom. For the random effects, var is the random effects variance component. Significant values are shown in bold.

We did, however, find benefits of a higher intensity and frequency hardening treatment (triple 37 °C) compared to a lower intensity and frequency hardening treatment (single 35 °C) under predation by spiders. The triple 37 °C hardening treatment therefore increased heat resistance and survival under predation by spiders compared to the single 35 °C hardening treatment. Cross-resistance, where exposure to one stressor enhances resistance to other stressors, has been found in association with some climatic stressors (Hoffmann et al., 2003). Predation risk and climatic factors may influence the same traits of an organism's biology such as foraging or metabolic rate (Cossins & Bowler, 1987; Rovero et al., 1999; Sanford, 1999; Trussell & Smith, 2000; Hawlena & Schmitz, 2010; Trussell & Schmitz, 2012). A potential explanation for our results is that heat hardening reduces the metabolic rate and/or the activity level. Reduced metabolic rate, together with accumulation of energy reserves, has been suggested as a general mechanism for stress resistance (Hoffmann & Parsons, 1989; Bubliy et al., 2012). Reduced activity is also thought to be one of the most efficient antipredator defence that reduces the encounter rate with predators, decreases detection by predators and increases time spent hiding (Werner & Anholt, 1993; Stoks et al., 2003, 2005). In addition, predation risk can alter metabolic rates (Rovero et al., 1999; Beckerman et al., 2007; Slos & Stoks, 2008) and elevate stress proteins (Kagawa & Mugiya, 2002; Pauwels et al., 2005). The heat hardening effects on fly physiology might therefore involve similar physiological responses as predation risk and decrease vulnerability to predation. Although these responses are beneficial under predation, they might still come with long-term costs, as a decreased activity also reduces foraging and usually results in reduced growth and fecundity (Werner & Anholt, 1993; Brodin & Johansson, 2004).

# Costs of extreme heat stress under predation pressure

Our results showed that flies surviving a 39 °C selection cycle had a lower survival under predation than control flies. In comparison with the hardening treatments, the selection cycle exposed the flies to a much more severe heat stress which very likely causes cellular and physiological damage. Heat shock has deleterious effect on the internal organization of the cell beyond unfolding of proteins (Sørensen et al., 2003). With increasing temperature and a longer heat exposure than experienced under the hardening treatment, the damage is likely to increase and benefits decrease. These damages are likely to affect the flies' physiological performance and to impair the detection and escape of the predators. Benefits of short-term heat hardening on predation can therefore only be expected as long as the heat hardening is not too harsh (Angilletta, 2009).

# Frequency- and intensity-dependent heat hardening effects under predation pressure

We only found significant differences between single 35 °C and triple 37 °C hardening treatment, but not between the control and the hardening treatments in survival under predation by spiders. In addition, we found costs of severe heat exposure (39 °C) under predation pressure by spiders and mantids. These results may suggest that the relationship between heat exposure and predation avoidance is nonlinear. One possible scenario could be that metabolic rate (or activity) has an inverse U-shaped reaction norm across different levels of heat exposure. The mild single 35 °C treatment might increase metabolic rate and might therefore be slightly costly under predation risk compared to the control treatment. The moderate triple 37 °C hardening treatment becomes beneficial under predation, due to a decreased metabolic rate (or activity). An extreme heat stress, such as the 39 °C exposure at generation 0, becomes costly again, which might be due to a strongly decreased metabolic rate (or activity) and/or due to its deleterious effects on cell functioning as mentioned above. There is some support for this scenario: first, metabolic rates are assumed to increase with temperature up to an optimal temperature in most insects (Angilletta, 2009). In addition, evidence for a decreased metabolic rate after an extreme heat stress has been found in damselflies (Dinh *et al.*, 2016). Whether metabolic rate (or activity) indeed follows an inverse U-shaped reaction norm across different levels of heat stress remains to be tested.

# Sex-specific predation avoidance after heat exposure

Our results also revealed some sex effects in predation avoidance after heat exposure. Whereas the hardening effects on survival under predation by spiders were similar in both sexes, we found that the sexes responded differently to the hardening treatments under predation by mantids. In addition, only the males had reduced survival under predation by mantids after the 39 °C selection cycle at generation 0. What is driving sex-specific predation avoidance is currently not known, but one explanation could be that the reaction norms of metabolic rate (or activity) differ between the sexes across different levels of heat exposure, which remains to be tested.

# No evidence for a genetic trade-off between heat resistance and predation avoidance

Little is known about predation of natural *D. me-lanogaster* populations. Insects are generally predated by a wide range of insects, as well as other species, such as vertebrates and birds which can play a significant role in insect population dynamics (Speight *et al.*, 2008). Predation pressure can vary in space and time and can have severe effects on population demography (Speight *et al.*, 2008) as evident from classic life table studies such as those undertaken on the population demography of winter moths (Varley & Gradwell, 1960).

At this stage, we have no evidence that selection for increased heat resistance decreases survival under predation by jumping spiders or mantids. In addition, both sets of lines selected for decreased predation risk (episodic and continuous predation) did not differ in heat resistance. These experiments suggest that there is no strong genetic covariance between predation avoidance and heat resistance in *D. melanogaster* and suggest that heat adaptation is not limited by biotic interactions associated with predation. Very few studies have tested for biotic costs of evolved stress resistance. Studies on the water flea have found that the evolution of increased pesticide resistance has costs under predation risk (Jansen *et al.*, 2011). However, pesticide resistance is based on different mechanisms than thermal resistance. Currently, we have limited knowledge on genetic constraints associated with predation and their impact on climate adaption, but the results presented here suggest that the observed phenotypic patterns (in terms of hardening effects and stress costs) are not reflected in evolutionary changes. Any impacts of predation costs and benefits on natural population are likely to be complex, as not only prey species but also predators are exposed to heat stress and predator species may respond differently to such stress (Harmon *et al.*, 2009).

### Acknowledgments

We thank Ronald Lee for collecting the flies and Lea Rako, Jennifer Shirriffs, Fernando Diaz, Kelly Richardson, Anjali Goundar and Yoshinori Endo for support with rearing of the laboratory lines. This research was funded by the Swiss National Science Foundation (PBEZP3\_140043 and PA00P3\_145372 to SH) and by the Australian Research Council and the CSIRO Science Industry Endowment Fund (to AAH) and by a National Science and Engineering Research Council of Canada Discovery award (to ID). The authors declare no conflict of interest.

### References

- Alton, L.A., Wilson, R.S. & Franklin, C.E. 2010. Risk of predation enhances the lethal effects of UV-B in amphibians. *Glob. Change Biol.* 16: 538–545.
- Angilletta, M.J. 2009. Thermal Adaptation: A Theoretical and Empirical Synthesis. Oxford University Press, Oxford.
- Barry, M.J. 2000. Effects of endosulfan on Chaoborus-induced life-history shifts and morphological defenses in *Daphnia pulex*. J. Plankton Res. **22**: 1705–1718.
- Beckerman, A.P., Wieski, K. & Baird, D.J. 2007. Behavioural versus physiological mediation of life history under predation risk. *Oecologia* 152: 335–343.
- Blumstein, D., Lea, A., Olson, L. & Martin, J. 2010. Heritability of anti-predatory traits: vigilance and locomotor performance in marmots. J. Evol. Biol. 23: 879–887.
- Brodin, T. & Johansson, F. 2004. Conflicting selection pressures on the growth/predation-risk trade-off in a damselfly. *Ecology* 85: 2927–2932.
- Brokordt, K., Farías, W., Lhorente, J.P. & Winkler, F. 2012. Heritability and genetic correlations of escape behaviours in juvenile scallop *Argopecten purpuratus*. *Anim. Behav.* 84: 479– 484.
- Bubliy, O.A. & Loeschcke, V. 2005. Correlated responses to selection for stress resistance and longevity in a laboratory population of *Drosophila melanogaster*. J. Evol. Biol. 18: 789– 803.
- Bubliy, O.A., Kristensen, T.N., Kellermann, V. & Loeschcke, V. 2012. Plastic responses to four environmental stresses and

cross-resistance in a laboratory population of *Drosophila mela-nogaster*. Funct. Ecol. **26**: 245–253.

- Bubliy, O.A., Kristensen, T.N. & Loeschcke, V. 2013. Stressinduced plastic responses in *Drosophila simulans* following exposure to combinations of temperature and humidity levels. *J. Exp. Biol.* **216**: 4601–4607.
- Charmantier, A., McCleery, R.H., Cole, L.R., Perrins, C., Kruuk, L.E. & Sheldon, B.C. 2008. Adaptive phenotypic plasticity in response to climate change in a wild bird population. *Science* **320**: 800–803.
- Chevin, L.M., Collins, S. & Lefèvre, F. 2013. Phenotypic plasticity and evolutionary demographic responses to climate change: taking theory out to the field. *Funct. Ecol.* **27**: 967–979.
- Cossins, A.R. & Bowler, K. 1987. Temperature Biology of Animals. Chapman and Hall, New York, NY, USA.
- DeNieu, M., Pitchers, W. & Dworkin, I. 2014. Adaptation to a novel predator in *Drosophila melanogaster*: How well are we able to predict evolutionary responses? bioRxiv:005322.
- Dinh, K., Janssens, L. & Stoks, R. 2016. Exposure to a heat wave under food limitation makes an agricultural insecticide lethal: a mechanistic laboratory experiment. *Glob. Change Biol.* 22: 3361–3372.
- Egea-Serrano, A., Hangartner, S., Laurila, A. & Räsänen, K. 2014. Multifarious selection through environmental change: acidity and predator-mediated adaptive divergence in the moor frog (*Rana arvalis*). *Proc. R. Soc. B* 281: 20133266.
- Eränen, J.K., Nilsen, J., Zverev, V.E. & Kozlov, M.V. 2009. Mountain birch under multiple stressors - heavy metal-resistant populations co-resistant to biotic stress but maladapted to abiotic stress. *J. Evol. Biol.* **22**: 840–851.
- Feder, J.H., Rossi, J.M., Solomon, J., Solomon, N. & Lindquist, S. 1992. The consequences of expressing hsp70 in *Drosophila* cells at normal temperatures. *Genes Dev.* 6: 1402–1413.
- Gelperin, A. 1968. Feeding behaviour of the praying mantis: a learned modification. *Nature* **219**: 399–400.
- Ghalambor, C.K., Reznick, D.N. & Walker, J.A. 2004. Constraints on adaptive evolution: the functional trade-off between reproduction and fast-start swimming performance in the Trinidadian guppy (*Poecilia reticulata*). Am. Nat. 164: 38–50.
- Gilchrist, G.W. & Huey, R.B. 1999. The direct response of *Drosophila melanogaster* to selection on knockdown temperature. *Heredity* **83**: 15–29.
- Grant, B. & Mettler, L. 1969. Disruptive and stabilizing selection on the" escape" behavior of *Drosophila melanogaster*. *Genetics* **62**: 625–637.
- Hanazato, T. 2001. Pesticide effects on freshwater zooplankton: an ecological perspective. *Environ. Pollut.* **112**: 1–10.
- Hangartner, S. & Hoffmann, A.A. 2016. Evolutionary potential of multiple measures of upper thermal tolerance in *Drosophila melanogaster. Funct. Ecol.* **30**: 442–452.
- Harley, C.D. 2011. Climate change, keystone predation, and biodiversity loss. *Science* **334**: 1124–1127.
- Harmon, J.P., Moran, N.A. & Ives, A.R. 2009. Species response to environmental change: impacts of food web interactions and evolution. *Science* **323**: 1347–1350.
- Hawlena, D. & Schmitz, O.J. 2010. Herbivore physiological response to predation risk and implications for ecosystem nutrient dynamics. *Proc. Natl. Acad. Sci. USA* 107: 15503– 15507.
- Hercus, M.J., Loeschcke, V. & Rattan, S.I. 2003. Lifespan extension of *Drosophila melanogaster* through hormesis by repeated mild heat stress. *Biogerontology* **4**: 149–156.

- Hoffmann, A.A. & Parsons, P.A. 1989. An integrated approach to environmental-stress tolerance and life-history variation desiccation tolerance in *Drosophila*. *Biol. J. Linn. Soc.* 37: 117– 136.
- Hoffmann, A.A. & Parsons, P.A. 1991. Evolutionary Genetics and Environmental Stress. Oxford Univ. Press, Oxford, UK.
- Hoffmann, A.A., Anderson, A. & Hallas, R. 2002. Opposing clines for high and low temperature resistance in *Drosophila melanogaster*. Ecol. Lett. 5: 614–618.
- Hoffmann, A.A., Sørensen, J.G. & Loeschcke, V. 2003. Adaptation of *Drosophila* to temperature extremes: bringing together quantitative and molecular approaches. *J. Therm. Biol* 28: 175–216.
- Hoffmann, A.A., Chown, S.L. & Clusella-Trullas, S. 2013. Upper thermal limits in terrestrial ectotherms: how constrained are they? *Funct. Ecol.* **27**: 934–949.
- Huang, L.-H., Chen, B. & Kang, L. 2007. Impact of mild temperature hardening on thermotolerance, fecundity, and Hsp gene expression in *Liriomyza huidobrensis*. J. Insect Physiol. 53: 1199–1205.
- Huber, H., Kane, N.C., Heschel, M.S., von Wettberg, E.J., Banta, J., Leuck, A.M. *et al.* 2004. Frequency and microenvironmental pattern of selection on plastic shade-avoidance traits in a natural population of *Impatiens capensis*. *Am. Nat.* 163: 548–563.
- Huey, R.B. & Kingsolver, J.G. 1993. Evolution of resistance to high temperature in ectotherms. *Am. Nat.* **142**: 21–46.
- IPCC. 2014. Summary for Policymakers. In: Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A: Global and Sectoral Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change.
- Jackson, R. & Pollard, S. 1996. Predatory behavior of jumping spiders. Annu. Rev. Entomol. 41: 287–308.
- Jansen, M., De Meester, L., Cielen, A., Buser, C.C. & Stoks, R. 2011. The interplay of past and current stress exposure on the water flea *Daphnia*. *Funct. Ecol.* **25**: 974–982.
- Kagawa, N. & Mugiya, Y. 2002. Brain HSP70 mRNA expression is linked with plasma cortisol levels in goldfish (*Carassius auratus*) exposed to a potential predator. *Zoolog. Sci.* 19: 735–740.
- Karl, I., Becker, M., Hinzke, T., Mielke, M., Schiffler, M. & Fischer, K. 2014. Interactive effects of acclimation temperature and short-term stress exposure on resistance traits in the butterfly *Bicyclus anynana*. *Physiol. Entomol.* **39**: 222–228.
- Kingsolver, J., Massie, K., Ragland, G. & Smith, M. 2007. Rapid population divergence in thermal reaction norms for an invading species: breaking the temperature–size rule. *J. Evol. Biol.* **20**: 892–900.
- Krebs, R. & Loeschcke, V. 1994. Costs and benefits of activation of the heat-shock response in *Drosophila melanogaster*. *Funct. Ecol.* 8: 730–737.
- Kristensen, T.N., Hoffmann, A.A., Overgaard, J., Sørensen, J.G., Hallas, R. & Loeschcke, V. 2008. Costs and benefits of cold acclimation in field-released *Drosophila*. *Proc. Natl. Acad. Sci. USA* **105**: 216–221.
- Littell, R.C., Milliken, G.A., Stroup, R.D., Wolfinger, R.D. & Schabenberger, O. 2006. *SAS for Mixed Models*, 2nd edn. SAS Institute, Cary, NC, USA.
- Loeschcke, V. & Hoffmann, A.A. 2007. Consequences of heat hardening on a field fitness component in *Drosophila* depend on environmental temperature. *Am. Nat.* **169**: 175–183.

<sup>© 2017</sup> EUROPEAN SOCIETY FOR EVOLUTIONARY BIOLOGY. J. EVOL. BIOL. 30 (2017) 1153–1164 JOURNAL OF EVOLUTIONARY BIOLOGY © 2017 EUROPEAN SOCIETY FOR EVOLUTIONARY BIOLOGY

- McColl, G., Hoffmann, A.A. & McKechnie, S.W. 1996. Response of two heat shock genes to selection for knockdown heat resistance in *Drosophila melanogaster*. *Genetics* 143: 1615–1627.
- Miller, L.P., Matassa, C.M. & Trussell, G.C. 2014. Climate change enhances the negative effects of predation risk on an intermediate consumer. *Glob. Change Biol.* 20: 3834–3844.
- Mousseau, T.A. & Fox, C.W. 1998. The adaptive significance of maternal effects. *Trends Ecol. Evol.* 13: 403–407.
- Pauwels, K., Stoks, R. & De Meester, L. 2005. Coping with predator stress: interclonal differences in induction of heatshock proteins in the water flea *Daphnia magna. J. Evol. Biol.* 18: 867–872.
- Relyea, R.A. 2004. Growth and survival of five amphibian species exposed to combinations of pesticides. *Environ. Toxicol. Chem.* **23**: 1737–1742.
- Relyea, R.A. 2005. The heritability of inducible defenses in tadpoles. *J. Evol. Biol.* **18**: 856–866.
- Relyea, R.A. & Mills, N. 2001. Predator-induced stress makes the pesticide carbaryl more deadly to gray treefrog tadpoles (*Hyla versicolor*). *Proc. Natl. Acad. Sci. USA* **98**: 2491–2496.
- Rogell, B., Hofman, M., Eklund, M., Laurila, A. & Höglund, J. 2009. The interaction of multiple environmental stressors affects adaptation to a novel habitat in the natterjack toad *Bufo calamita. J. Evol. Biol.* 22: 2267–2277.
- Rovero, F., Hughes, R.N. & Chelazzi, G. 1999. Cardiac and behavioural responses of mussels to risk of predation by dogwhelks. *Anim. Behav.* **58**: 707–714.
- Sanford, E. 1999. Regulation of keystone predation by small changes in ocean temperature. *Science* **283**: 2095–2097.
- Scheiner, S.M. 1993. Genetics and evolution of phenotypic plasticity. *Annu. Rev. Ecol. Evol. Syst.* 24: 35–68.
- Schiffer, M., Hangartner, S. & Hoffmann, A.A. 2013. Assessing the relative importance of environmental effects, carry-over effects and species differences in thermal stress resistance: a comparison of Drosophilids across field and laboratory generations. J. Exp. Biol. 216: 3790–3798.
- Schou, M., Kristensen, T.N., Kellermann, V., Schlötterer, C. & Loeschcke, V. 2014. A *Drosophila* laboratory evolution experiment points to low evolutionary potential under increased temperatures likely to be experienced in the future. *J. Evol. Biol.* 27: 1859–1868.
- Schulte, P.M. 2007. Responses to environmental stressors in an estuarine fish: Interacting stressors and the impacts of local adaptation. J. Therm. Biol 32: 152–161.
- Sinclair, B.J., Vernon, P., Klok, C.J. & Chown, S.L. 2003. Insects at low temperatures: an ecological perspective. *Trends Ecol. Evol.* 18: 257–262.
- Slos, S. & Stoks, R. 2008. Predation risk induces stress proteins and reduces antioxidant defense. *Funct. Ecol.* 22: 637–642.
- Sørensen, J., Kristensen, T.N. & Loeschcke, V. 2003. The evolutionary and ecological role of heat shock proteins. *Ecol. Lett.* 6: 1025–1037.
- Sørensen, J., Kristensen, T.N. & Overgaard, J. 2016. Evolutionary and ecological patterns of thermal acclimation capacity in *Drosophila*: is it important for keeping up with climate change? *Curr. Opin. Insect Sci.* **17**: 98–104.
- Speight, M.R., Hunter, M.D. & Watt, A.D. 2008. Ecology of Insects: Concepts and Applications. Blackwell Science Ltd., Oxford, UK.

- Stirling, D., Réale, D. & Roff, D. 2002. Selection, structure and the heritability of behaviour. J. Evol. Biol. 15: 277–289.
- Stoks, R., McPeek, M., Mitchell, J. & Crespi, B. 2003. Evolution of prey behavior in response to changes in predation regime: damselflies in fish and dragonfly lakes. *Evolution* 57: 574–585.
- Stoks, R., De Block, M., Van de Meutter, F. & Johansson, F. 2005. Predation cost of rapid growth: behavioural coupling and physiological decoupling. J. Anim. Ecol. 74: 708–715.
- Teplitsky, C., Piha, H., Laurila, A. & Merilä, J. 2005. Common pesticide increases costs of antipredator defenses in *Rana temporaria* tadpoles. *Environ. Sci. Technol.* **39**: 6079–6085.
- Teplitsky, C., Räsänen, K. & Laurila, A. 2007. Adaptive plasticity in stressful environments: acidity constrains inducible defences in *Rana arvalis. Evol. Ecol. Res.* **9**: 447–458.
- Trussell, G. & Schmitz, O. 2012. Species functional traits, trophic control and the ecosystem consequences of adaptive foraging in the middle of food chains. In: *Trait-Mediated Indirect Interactions: Ecological and Evolutionary Perspectives* (T. Ohgushi, O. Schmitz & R.D. Holt, eds), pp. 324–338. Cambridge University Press, Cambridge, UK.
- Trussell, G.C. & Smith, L.D. 2000. Induced defenses in response to an invading crab predator: an explanation of historical and geographic phenotypic change. *Proc. Natl. Acad. Sci. USA* 97: 2123–2127.
- Varley, G. & Gradwell, G. 1960. Key factors in population studies. J. Anim. Ecol. 29: 399–401.
- Walsh, B. & Blows, M.W. 2009. Abundant genetic variation+ strong selection= multivariate genetic constraints: a geometric view of adaptation. *Annu. Rev. Ecol. Evol. Syst.* 40: 41–59.
- Werner, E.E. & Anholt, B.R. 1993. Ecological consequences of the trade-off between growth and mortality rates mediated by foraging activity. *Am. Nat.* **142**: 242–272.
- Willett, C.S. 2010. Potential fitness trade-offs for thermal tolerance in intertidal copepod *Tigriopus californicus*. *Evolution* **64**: 2521–2534.

### **Supporting information**

Additional Supporting Information may be found online in the supporting information tab for this article: **Table S1** Generalized linear mixed model of binary survival (yes or no) of female and male *D. melanogaster* under predation pressure by (A) jumping spiders and (B) juvenile mantids.

**Table S2** Generalized linear mixed model of binary survival (yes or no) of female and male *D. melanogaster* under predation pressure by (A) a jumping spider and (B) a juvenile mantids.

**Data S1** Description of the episodic and continuous predation lines.

Data deposited at Dryad: https://doi.org/10.5061/dryad.sg8c5

Received 30 November 2016; revised 31 March 2017; accepted 31 March 2017